

Replacement of carbon sources and determination of butyric acid

- 1. Culture of *Clostridium tyrobutyricum* engineered bacteria Ct (Pthl F/Xpk-BD) using TGY medium supplemented with different carbon sources**
- 2. Fermentation broths of Ct (Pthl F/Xpk-BD) in media containing different carbon sources were collected and assayed for product content using HPLC**
- 3. Analysis of HPLC data**

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Recorder:Yijiu Lu

7/8/2023 Monday

Steps-*Clostridium tyrobutyricum* activation:

Medium preparation and sterilization: on the basis of TGY medium, glucose was replaced with equimolar amounts of fructose and xylose, respectively, 50mL system

***Clostridium tyrobutyricum* activation:** 1 mL *Ct (Pthl F/Xpk-BD)*, no resistance added, add to pre-prepared TGY medium and incubated for 12 hours.



8/8/2023 Tuesday

Steps-*Clostridium tyrobutyricum* secondary activation:

***Ct(Pthl F/Xpk-BD)*, add double antibody (Tm and Dcyc),50mL, 12 hours**

Inoculation: the activated bacteria were inoculated into the medium with different carbon sources; the initial medium was taken and samples were retained

9/8/2023 Wednesday

Steps-take the samples:

Samples of the broth were taken once every 6-8 hours and OD₆₀₀ was measured at the same time.

10/8/2023 Thursday

Steps-Product Concentration Detection:

Liquid mobile phase: 267 μ L of concentrated sulfuric acid in 1L of water, filtration, ultrasonication.

Sample processing: centrifugation, membrane transfer, sample loaded to HPLC



11/8/2023 Friday

Steps-Record HPLC results:

Recording of results from HPLC

14/8/2023 Monday

Steps-Solution preparation:

1. hydroxylamine solution: 4M hydroxylamine hydrochloride: 3.5M NaOH=1:1,v/v
2. ferric chloride solution: 5% ferric chloride: 12% trichloroacetic acid: 3M HCl=1:1:1,v/v/v

***Clostridium tyrobutyricum* activation:** Control; *Ct(Pthl F/Xpk-BD)*; *Ct(Pthl F/Xpk-QS)*, no resistance, 1mL, 12 hours

15/8/2023 Tuesday

Steps-*Clostridium tyrobutyricum* secondary activation:

Control; *Ct(Pthl F/Xpk-BD)*; *Ct(Pthl F/Xpk-QS)*, add double antibody (Tm and Dcyc), 50mL, 12 hours

Medium preparation and sterilization:

on the basis of TGY medium, replace glucose with equal molar amounts of fructose and xylose respectively, 50mL system

Inoculation: the activated bacteria were inserted into the medium.

16/8/2023 Wednesday

Steps-Measuring acetyl phosphate:

1. Take the bacteria in the pre-stabilization period, wash them twice with PBS (10mM, pH 7.2-7.4), and ultrasonically break them;
2. Centrifuge the broken cells (12000rpm,5min), and set the supernatant on ice;
3. Take 500 μ L of supernatant, add 250 μ L of hydroxylamine solution for reaction, and leave it at room temperature for 30 min;
4. Add 750 μ L ferric chloride solution to develop the color, and leave it at room temperature for 10min, avoiding light;
5. Determine the absorbance value at 540nm.

The data are as follows

	504	540	505	540
0	0.178	0.043	0.111	0.100
3	0.974	0.726	0.824	0.757
6	0.938x2	0.695x2	1.914 (0.75x2)	1.297 (0.66x2)
9	0.880x3	0.646x3	0.704x3	0.444x3
12	0.646x4	0.875x4	1.027x3	0.920x3
15	1.187x4	0.865x4	1.223x4	1.078x4
K	0.854	0.826	0.142	0.676
	0.816	0.414		0.637
	0.791	0.621		0.575
BD	0.949	0.740	0.222	0.559
	0.967	0.552		0.753
	0.956	0.653		0.739
BS	0.944	0.653	0.142	0.778
	0.927	0.340		0.707
	0.849	0.745		0.774
DF	0.922			0.694
				0.703
				0.622
				2.492
				2.312
				2.238
				2.135
				2.324
				2.922
				2.881
				2.529
				2.571